Update on Pediatric Cancer Surveillance Recommendations for Patients with

Neurofibromatosis Type 1, Noonan Syndrome, CBL Syndrome, Costello Syndrome, and

Related RASopathies

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Abstract:

Neurofibromatosis type 1 (NF1), Noonan syndrome and related syndromes, grouped as the RASopathies, result from dysregulation of the RAS-MAPK pathway and demonstrate varied multisystemic clinical phenotypes. Together the RASopathies are among the more prevalent genetic cancer predisposition syndromes and require nuanced clinical management. When compared to the general population, children with RASopathies are at significantly increased risk of benign and malignant neoplasms. In the last decade, clinical trials have shown that targeted therapies can improve outcomes for low-grade and benign neoplastic lesions but have their own challenges, highlighting the multi-disciplinary care needed for such individuals, specifically those with NF1. This perspective, which originated from the 2023 AACR Childhood Cancer Predisposition Workshop, serves to update pediatric oncologists, neurologists, geneticists, counselors, and other healthcare professionals on revised diagnostic criteria, review previously published surveillance guidelines, and harmonize updated surveillance recommend to the general population, children with RASopathies are at significantly increased

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Introduction:

Neurofibromatosis type 1 (NF1), Noonan syndrome (NS) and related RASopathies, including Costello syndrome (CS), Cardiofaciocutaneous (CFC) syndrome and Legius syndrome (LS) are genetic conditions caused by dysregulation of the RAS-mitogen activated protein kinase (MAPK) pathway. People with these germline genetic changes face variable neurologic, cardiac, skeletal, and other medical challenges from an early age. With the exception of LS, children with RASopathies are at significantly increased cancer risk compared to the general population(1-6). Advances in genotype-phenotype correlation have led to diagnostic updates and improved management of individuals with these syndromes. Molecular therapeutics inhibiting the RAS-MAPK pathway now expand the treatment options for patients with NF1 and symptomatic, unresectable plexiform neurofibroma or glioma, which is promising to reduce and prevent severe tumor burden and subsequent morbidity(7-9). As diagnostic and treatment modalities evolve, surveillance recommendations must be amended to optimize patient care and stay true to the pillars of surveillance: early tumor detection with potential for intervention while minimizing the risks of surveillance when possible(4,10-15). In this effort, we present updated consensus cancer surveillance recommendations for patients with NF1, NS, CS, and other RASopathies.

Neurofibromatosis Type 1

NF1 incidence, genetic etiology, and updates to the clinical criteria:

NF1 is among the most common cancer predisposition syndromes (CPS) and has a birth prevalence of 1/2,000-1/3,000 persons(16). It is an autosomal dominant syndrome resulting from pathogenic variants (PV) in the gene *NF1*, encoding neurofibromin, a key negative regulator in the RAS-MAPK pathway.

Approximately half of NF1 cases occur *de novo* and penetrance is generally complete with variable expressivity(17). Somatic mosaicism is well-documented in NF1, and mosaic individuals may have variable to few clinical features(18-20). Genotype-phenotype associations have been described, but for the majority of variants, the expressivity cannot be predicted based on the specific variant(20-22). Currently, there is no recommendation to change NF1 tumor surveillance recommendations in the setting of mosaicism or for genotypes with stronger phenotype correlation.

In 2021, Legius et al., revised the diagnostic criteria for NF1, with a specific focus on differentiating it from Legius syndrome, a RASopathy caused by *SPRED1* PV, which features the most phenotypic overlap with NF1(23). This clinical update represented the first significant alteration to the clinical diagnostic criteria since they were introduced in 1987(24). Moreover, when a patient is suspected to have NF1 but does not meet full clinical criteria and has negative *NF1* testing, other conditions with overlapping features such as Legius syndrome (*SPRED1*)*,* other (mosaic) RASopathies (e.g. *KRAS), CDKN2A*-Related Melanoma-Astrocytoma Syndrome (*CDKN2A*)*,* and constitutional mismatch repair deficiency (*PMS2, MLH1, MSH2, MSH6*) should be considered.

Tumor Risk and Natural history in NF1

In individuals with NF1, most neoplasms involve the nervous system, including gliomas, benign neurofibromas, borderline and malignant peripheral nerve sheath tumors (MPNST). Furthermore, *NF1*-associated tumors are age-dependent. Optic pathway gliomas (OPG)

present early in childhood. Other cancers, including juvenile myelomonocytic leukemia (JMML), rhabdomyosarcoma (RMS), and neuroblastoma (NB), though rare, can develop early in childhood at higher frequencies than in the general population. Plexiform neurofibromas (PN) are likely congenital but grow in the first two decades of life, and atypical neurofibromatous neoplasms of uncertain biologic potential (ANNUBP) are pre-malignant tumors that typically develop within, or nearby, a known PN in the second to third decades of life(25-27). Meanwhile, MPNST, pheochromocytomas, gastrointestinal stromal tumors (GIST), and breast cancers usually develop in adults, though many are diagnosed at significantly younger ages than in the general population(6).

Central Nervous System Tumors

The most common central nervous system tumor type in NF1 is low-grade glioma (LGG; ~20%), frequently affecting the optic pathway (OPG) and presenting at age <8 years (median: 4-5 years). The vast majority of NF1-related OPGs will not progress after their initial diagnosis, and may follow an indolent course with the potential for spontaneous growth arrest. However, 15-20% of these tumors progress, result in visual deterioration, strabismus, proptosis, papilledema, and nystagmus, and require intervention. Patients with radiological involvement of the posterior optic tracts, age <2 years, and of female sex may be at increased risk of progression(28,29). Irradiation is usually avoided in NF1-associated gliomas due to vascular and other complications. Chemotherapy is traditionally the mainstay of therapy though response to targeted therapies including MEK-inhibitors is promising with multiple studies neurontocous neoplasms of uncertain biologic potential (ANNUBP) are pre-mailgnant
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Pilocytic and diffuse astrocytomas may occur as focal brainstem enlargement in <10% of individuals with NF1 37 . Mean age of presentation is 7 years and <5% tumors identified require treatment(32) (33). Treatment may ultimately be necessary in patients with neurological deterioration and a biopsy may be required prior to treatment as these lesions have the potential to transform from low to high-grade.

Malignant transformation of pre-existing LGG is rare in NF1, but well-recognized. Anaplastic transformation of some LGG may be independent of radiation therapy. Such grade 3 and 4 diffuse astrocytomas do not occur in the optic pathway, but may develop in the hemispheres, thalamus, cerebellum, and spinal cord and harbor aberrations in *ATRX*, *TP53* and *CDKN2A/2B* (while lacking histone or *IDH* mutations)(34).

Peripheral Nerve Sheath Tumors

PNs are histologically benign tumors seen in ~50% of patients with NF1 and develop following somatic loss of the remaining wild-type allele in Schwann cells. These lesions are likely congenital, have faster growth in young children, and may cause pain, disfigurement, and compression symptoms. Clinically meaningful, and durable, benefit from MEK-inhibitors in younger children has led to FDA approval of selumetinib for children \geq age 2 years with symptomatic, inoperable PNs(7-9). Trametinib, another MEK-inhibitor, can be effective in the management of PNs where selumetinib is not available(35,36). In addition, the receptor tyrosine kinase inhibitor cabozantinib has shown activity in the treatment of patients with NF1 $PN \geq 16$ years old(37). It is not known whether the risk of transformation to ANNUBP or MPNST is modified by targeted therapies, mandating the need for close attention to clinical potential to transform from tow to nigh-grade.

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Peripheral nerve sheath tumors with atypical growth patterns, symptoms, or imaging characteristics from an underlying PN should raise suspicion for ANNUBP. These tumors are thought to be pre-malignant lesions, although the timeline for malignant transformation remains unclear. They are often characterized by magnetic resonance imaging (MRI) findings of distinct nodular lesions (DNL), a >3 cm nodule within or nearby a known PN with loss of a central "target sign", and grow faster than the surrounding PN(26,27,39,40). ANNUBP are defined by histological features and genomic studies identified additional biallelic somatic loss of *CDKN2A/B* (25). While *TP53* and PRC2 genes including *EED* and *SUZ12* are common variants seen in MPNST, these have not been identified in ANNUBP to date. ANNUBP should be considered for surgery with gross total resection using a nerve-sparing approach without wide-margins at a center with surgeons familiar with NF1 whenever feasible(41-43).

Full malignant transformation of PN into MPNST occurs in 8-16% of NF1 patients. This is rare in childhood and peak incidence is in the 3rd-4th decade. Risk factors for MPNST development in individuals with NF1 include those with microdeletion involving *SUZ12*, *NF1* missense variants affecting codons 844-848, previous ANNUBP, neuropathy, previous radiation, and an NF1-relative with MPNST(44-49). In individuals with a known PN, rapid growth of a DNL, intra-tumoral cystic changes, and evidence of necrosis on imaging are concerning for 'high-risk' pathological features, and combined use of MRI- and 18Ffluorodeoxyglucose positron-emission (18F-FDG PET)-based imaging may facilitate accurate and timely diagnosis of MPNST. Primary resection with wide negative margins, if feasible, is strongly recommended(50).

Other malignancies and young adult considerations

While a diagnosis of NF1 is enriched among JMML patients, the estimated risk in patients with NF1 is <1%(11). Fusion-negative embryonal RMS has an overall risk of <1% but is enriched in males and demonstrates a predilection for the genitourinary system(51-54). Clinical education should be provided to patients and families regarding the increased risk of these tumors, but surveillance is not indicated.

While the focus of these recommendations is for childhood surveillance, it is important to recognize increased adult malignancies. Many individuals are not the first in their family with the cancer predisposition syndrome highlighting the need for awareness and family care. Further, preparation for the transition into adulthood has many nuances, but adult surveillance is vital for adolescents to understand prior to their transition from pediatrics. Detailed adult surveillance recommendations have been previously published(55). Specifically, individuals with NF1 being born female are at increased risk for young-onset breast cancer (56). Breast cancer screening with mammography and consideration of breast MRI should be discussed between starting around age 30 years(55,57). Small imaging studies suggest that females with NF1 may have greater breast density and breast MRI, when available, should be incorporated into screening when mammograms cannot be adequately interpreted(58). Surveillance for tumors other than breast cancer is not currently recommended in adulthood. Education regarding additional adult cancer risk should be considered for the adolescent individual. Pheochromocytomas and paragangliomas have been reported in 1–5% of patients with NF1, diagnosed at a median age of 40–50 years(59). In adults, undifferentiated pleomorphic sarcoma is seen at increased rates and with inferior outcomes compared to sporadic cases(6). Glomus tumors of the digits are small benign tumors that present with localized tenderness,

10

severe paroxysmal pain and sensitivity to cold(60). Patients with NF1 have a 200-fold increased risk of developing GISTs, which present around age 50 years and typically lack alterations in *KIT* or *PDGFRA*. Melanoma is reported in <1% of individuals with NF1 but has an overall higher association and inferior survival as compared to sporadic cases(6).

Proposed tumor surveillance updates for pediatric patients with Neurofibromatosis Type 1

OPG: In individuals with confirmed NF1, those with pending testing and a family history, or if NF1 is clinically suspected, ophthalmology surveillance should start at age 6-8 months (Table 1). An ophthalmologic exam by a trained pediatric ophthalmologist or neuro-ophthalmologist is recommended for age-appropriate, comprehensive evaluation with visual acuity and visual fields. Optical coherence tomography may be utilized if available at an experienced center but does not replace standard visual exams. Visual assessment continues annually, if exams remain normal, until age 8 years, then every other year to age 18 years (**Table 1**). If an exam is concerning for visual compromise, a MRI of the brain and orbits should be obtained and follow up for close visual monitoring should occur within 3 months. If a tumor is identified, but vision remains stable, close monitoring without therapeutic intervention is warranted. As the visual exam guides intervention and often can be completed without sedation, many groups have removed imaging from surveillance of asymptomatic patients for OPG(11,12,61,62). However, a highly trained pediatric ophthalmologist or neuro-ophthalmologist is not always accessible and if visual assessment is unreliable or inconsistent \geq age 2 years, an MRI should be obtained(11,63). Caution should be used in interpreting an MRI without visual changes to avoid increased surveillance imaging studies, sedation, and treatment in patients that may

11

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never become symptomatic. Importantly, a normal MRI does not abrogate the recommendation for continued visual exams.

PN: Surveillance for PN has typically not been recommended, as interventions for asymptomatic PN were contraindicated. With the success of, and increasing access to, MEKinhibitors, early treatment may be considered for patients at high risk of developing PN-related morbidity(7-9). In this regard, a clinical trial is currently underway to determine whether surveillance and treatment initiation for asymptomatic, but high-risk due to location, PN will be beneficial (NCT06188741). Outside of a clinical trial, imaging in asymptomatic patients is not yet warranted (**Table 1**). If imaging is obtained due to another cause and internal tumors are identified incidentally, referral should be made to a pediatric oncology center with expertise in the care of NF1 patients with PN. If future growth of the tumors may interfere with vital organ function or cause acute neurologic complications, further dedicated imaging may guide interventions.

ANNUBP/DNL: In general, if a patient has a biopsy-proven ANNUBP, the current recommendation is for resection with a narrow margin if this can be achieved without significant morbidity. Additional consensus is needed for the recommended monitoring for ANNUBP that are not resected, but these lesions should be considered at increased risk for malignant transformation. For new or increasing symptomatic lesions, or rapidly growing lesions in older adolescence, imaging should be completed and if a DNL is detected in the area of symptoms, even with classically 'benign' features on imaging, marginal resection should be considered if feasible without significant morbidity. If resection is not feasible, imageguided biopsy should be completed with subsequent somatic testing for *CDKN2A*/*2B* loss.

MPNST: Whole-body MRI (WBMRI) has been recommended after puberty, in late adolescence when benign PN growth rates slow considerably, as a baseline for overall tumor burden prior to adulthood, and this recommendation remains unchanged (**Table 1**)(26,55,64-67). This is used to stratify risk for development of MPNST and educate patients to seek early medical attention for any change in symptoms at known tumor locations. A PN tumor burden >300mL or DNL should prompt closer clinical follow-up and education regarding symptoms of malignant transformation(26,67,68). Caution should be taken for patients with high-risk genotypes including an *NF1* microdeletion involving *SUZ12*, patients with a high tumor burden of PN in childhood, and patients with a personal or family history of MPNST, ANNUBP, or DNL(11,13,44). 18F-FDG PET/CT should remain reserved for patients with lesions concerning for MPNST such as painful or growing tumors and can be combined with dedicated regional MRI, including diffusion weight imaging (DWI) to use apparent diffusion coefficient (ADC) mapping for increased specificity and sensitivity in determining the most likely area of malignant transformation to target for biopsy or resection(11,67). Circulating tumor, or plasma cell-free DNA methods are currently still under investigation but are hoped to play a pivotal role in surveillance and early detection for ANNUBP and MPNST in high-risk patients in the future. In fact, individuals with NF1 may serve as exceptional models for circulating tumor DNA surveillance as imaging continues to be difficult to determine malignancy risk with both high sensitivity and specificity, and the secondary genetic drivers of these tumors have been well characterized(25,40,69,70).

If targeted MEK-inhibitor therapy is started for any reason in a patient with NF1, consider a WBMRI if available, or similar thick section MRI limited to the neck, chest, abdomen and pelvis (NCAP-WBMRI) prior to initiation as a baseline to better understand and monitor potential non-target tumor response (**Table 1**). Additional work through the AACR-Cancer Predisposition Working Group highlights radiologic detail and variations of WBMRI and can be found in a collaborating manuscript.

Noonan syndrome and CBL syndrome

Noonan syndrome incidence, genetic etiology, and clinical spectrum

NS is an autosomal dominant (with the rare exception of recessive *LZTR1* or *SPRED2* variants) condition with an estimated prevalence of 1 in 1,000 to 2,500 live births(71,72). Knowledge of the molecular basis of NS has increasingly evolved with variants in *PTPN11, SOS1, RAF1, RIT1, LZTR1, KRAS, SOS2, NRAS, RRAS, RRAS2, MRAS*, and *SPRED2* currently associated with disease(73-75). Individuals with a PV in *PTPN11* represent nearly half of NS cases and, therefore, the majority of data on cancer incidence is based on *PTPN11* cases. Clinically, NS is characterized by facial dysmorphisms, short stature, developmental delays, congenital heart defects (most often pulmonary valve stenosis, typically with dysplasia), hypertrophic cardiomyopathy, and increased cancer risk^(76,77).

Cancer risk in Noonan syndrome

The relative childhood cancer risk in NS is estimated to be increased approximately 8-fold over the general population(5). Due to the low baseline cancer risk in children, the high relative risk leads to a moderate absolute cancer risk(5). The spectrum of cancer types is broad and includes myeloid and lymphoblastic leukemia, RMS, NB, and glioma, among others(2,4,5,78- 81).

Specific *PTPN11* variants are associated with the development of an often-self-limiting myeloproliferative disorder (MPD)(4,82,83). This MPD can undergo transformation into JMML, although – apart from somatic events such as monosomy 7 - strong predictors for which individuals' MPD will self-resolve, and which will transform, are not yet known(84).

CBL syndrome

CBL syndrome has features overlapping with NS, including growth and developmental delays, hypertrophic cardiomyopathy, and vasculitis. This syndrome is diagnosed in an individual with a germline PV in *CBL*. Patients with CBL syndrome have an increased risk for JMML, associated with loss of heterozygosity in the bone marrow. The severity of CBL-related JMML is variable with both aggressive cases and those resolving without treatment(85,86). Unique to CBL syndrome, a patient's clinical course may be complicated by arteritis(85).

Proposed tumor surveillance updates for Noonan syndrome and CBL syndrome

While some individuals with NS first present with JMML, many first present with lessaggressive MPD, and there is no evidence to predict which patients' disease will spontaneously resolve and which will have frank transformation into JMML. Similarly, CBL syndrome patients can present with a less clinically aggressive form of JMML, and clinical management now recommends watchful waiting(87). Nevertheless, all patients with suspected MPD should be evaluated and followed by a pediatric hematologist/oncologist. There is no evidence that in otherwise healthy children with NS or CBL syndrome, surveillance with complete blood counts (CBC) is beneficial. Therefore, routine CBC has been removed from

surveillance, in alignment with other CPS predisposing to leukemia (See related manuscript through the AACR-Cancer Predisposition Working Group specific to leukemia surveillance). Clinical exam remains the mainstay of early intervention, and all patients, regardless of genotype, are recommended to have close monitoring including physical examination for hepatosplenomegaly especially during infancy and early childhood (**Table 2**). The absolute cancer risk for non-hematologic malignancies is not sufficiently high to warrant radiologic or laboratory surveillance. However, education of patients and families on the signs and symptoms of RMS, gliomas, and NB can be considered. Finally, there has been no evidence supporting the notion that use of growth hormones increases the risk or rate of tumor growth. Therefore, in patients with NS, and other RASopathies, we do not advise against the use of growth hormone therapy, if indicated clinically without other contraindications. Outside of a clinical trial, a brain MRI is not recommended in an asymptomatic child prior to starting growth hormone therapy.

Costello syndrome

Costello syndrome incidence, genetic etiology, and clinical spectrum

CS is characterized by heterozygous disease-causing variants in *HRAS* and is the RASopathy with the highest risk for malignancy. With a lower birth prevalence, at 1:380,000 births, it is much less common than NF1(88). Despite being an autosomal dominant condition, most patients with CS have a *de novo* variant as individuals with CS rarely have biological children, making inheritance of this syndrome rare.

Tumor spectrum and natural history in Costello syndrome

CS has an estimated 13% risk (95% CI: 9-16) of malignancy by age 20 years(3). This is predominantly due to an increased risk for RMS, NB, and transitional cell carcinoma of the bladder (TCC). Due to the high absolute cancer risks during the pediatric age range (**Table 3**), surveillance is recommended for these malignancies in all patients with CS(3,5,89). Genotypephenotype cancer risk correlations showed that *HRAS* p.Gly12Cys (37.5% cancer incidence) and *HRAS* p.Gly12Asp (38.7%) have higher risks of malignancy compared to the more common *HRAS* p.Gly12Ser (9.2%)(3). However, as *HRAS* p.Gly12Ser is found in over 80% of CS patients, it is difficult to provide reliable cancer risk estimates for non-p.Gly12Ser variants. To our knowledge, there have been no malignancies reported in rare patients with CS due to *HRAS* non-p.Gly12 substitution variants(2,3,5,15,90). These possible genotype-cancer risk associations should be discussed with the parents/guardians of an affected child with CS but do not currently change recommended surveillance strategies.

Proposed tumor surveillance updates for Costello syndrome

Rhabdomyosarcoma

RMS is diagnosed in children with CS at similar young ages compared to sporadic RMS and has not been diagnosed after age 14 years. RMS is most often found in the abdomen/pelvis (80% of patients with CS and known tumor location)(3). While sporadic RMS also occurs in the head and neck region in 35% of cases and in the extremities in 13% of cases, this appears less frequently in CS (one patient each with: orbital, sphenoid, foot, and chest location)(89,91)[,](92) Therefore, imaging surveillance efforts focus on the abdomen and pelvis for early identification and rely on physical exam for tumors arising outside of the abdomen/pelvis(89,91). Abdominopelvic ultrasound (US) should be started as early as a

disease-causing *HRAS* variant is identified and continue until age 14 years (**Table 3**). Due to US techniques and potential limitations relating to body habitus, sensitivity may decrease in older children and a dedicated, regional MRI should be pursued if US is considered nondiagnostic, or in patients with symptoms. Due to comorbidities and potential increase in repeated sedation for individuals with CS, MRI is not recommended as first line for screening.

Neuroblastoma

Young patients with CS are also at higher risk to develop NB. The age of diagnosis and location of NB do not differ from the general population, although data are limited. Screening should begin as early as an *HRAS* variant is identified, and continue until age 6 years, then with decreasing frequency from 6-10 years (**Table 3**). Not specific to CS, over 80% of NB occur in the adrenal glands and paraspinal sympathetic chain. As such, abdominopelvic US is the primary method for surveillance(14,89,93). The remainder of NB diagnoses primarily occur in the thoracic region, however, given the low estimated absolute risk of thoracic NB of <1%, chest radiograph is not generally recommended. Urine catecholamines levels are not a useful surveillance method for patients with CS, as these individuals have variable and elevated levels at baseline(94).

Bladder carcinoma

TCC of the bladder has been seen in up to 2.2% of patients with CS and shows the highest relative cancer specific risk increase because it is rarely diagnosed in the general pediatric population(2,3). In contrast to RMS and NB, TCC is rarely seen in infancy and more often occurs after age 10 years with an elevated risk throughout adolescence and adulthood.

Surveillance for TCC remains challenging as sensitive measures, such as cystoscopy, are highly invasive and typically require in-hospital anesthesia. Surveillance cystoscopies could be considered for effective surveillance every 2 years starting at age 10, with a role for patient and family shared decision making due to the invasive nature of the procedure **(Table 3)**(95). Urinalysis (UA) has previously been recommended to evaluate for hematuria due to its minimally invasive nature, although data are limited on sensitivity and specificity of this method(14,89) (95). Less invasive testing, such as urine cytology, may be most effective but should be done in the context of a clinical trial to understand concerning features that would prompt further work-up.

Adult tumors: Aside from TCC, there is not an identified increased incidence for common adult malignancies. This may be due to early non-malignancy related deaths in patients with CS, and likely does not reflect true risk. Annual clinical exams remain important throughout life and cancer surveillance recommendations for the general population should be followed(90).

Other RASopathies and their associated cancer risk

NS with multiple lentigines (NSML), previously referred to as multiple lentigines syndrome or LEOPARD syndrome, is associated with distinct disease-causing variants in the *PTPN11* gene (the most common being Y279C and T468M), *RAF1, MAP2K1,* and *BRAF*(96-100)*.* Clinical features of NSML include hypertrophic cardiomyopathy, pulmonary valve stenosis, short stature, pectus deformity, and [dysmorphic](https://www.ncbi.nlm.nih.gov/books/n/gene/glossary/def-item/dysmorphic/) facial features including widely spaced eyes and ptosis. Lentigines appear predominantly around age 4-5 years as hypertrophic, black-brown

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macules occurring on the face, neck and upper trunk(101). Cancers reported in association with NSML include leukemia, neuroblastoma, and melanoma(2,102).

Cardiofaciocutaneous (CFC) syndrome is currently estimated to affect 1 in 800,000 newborns and is caused by germline disease-causing variants in *BRAF, MAP2K1*, *MAP2K2*, and more rarely *KRAS* and *YWHAZ(103).* Individuals with CFC syndrome have been reported with leukemia, lymphoma, RMS, and hepatoblastoma, however, whether an increased cancer incidence in CFC syndrome exists remains unknown given limited cases(2,104).

Legius syndrome (LS) is caused by PVs in *SPRED1* and was first identified in individuals meeting previous diagnostic criteria for NF1 in the absence of *NF1* PV(105). Pigmentary changes similar to those seen in NF1, including café au lait macules and skin fold freckling, are a common feature of LS. However, tumor risks including neurofibromas and optic gliomas are absent(106). While cancers have been reported in individuals with *SPRED1* PVs, it is unknown whether these were related to the *SPRED1* variants or occurred by chance(105,107,108). Given the phenotypic overlap between LS and NF1, it is important to differentiate the two conditions in individuals presenting with café au lait macules as it can have significant impact on screening recommendations(23).

Tumor surveillance in other RASopathies

We recommend that in patients with NSML and CFC syndrome, the surveillance guidelines for NS are followed (**Table 2**). In NSML, the skin should be examined at least annually, and a dermatologist should be involved if there are specific concerns(102). No cancer surveillance is recommended in LS.

Discussion and Future Directions:

Improved understanding of the genetic basis to these syndromes, genotype-phenotype correlations, and the natural history of at-risk cancers will continue to advance surveillance recommendations. Refining surveillance techniques is necessary to provide the most impact on improving patient outcomes while decreasing ineffective surveillance. Further, enrollment of these individuals in a cancer predisposition or disease-specific registry is recommended to gain the necessary data and advance recommendations. Shared decision making is needed between healthcare providers and families. As many of these tumors present in adolescence and young adulthood, a smooth transition of healthcare should be emphasized, and young adults are encouraged to take an active role in their health. Guidelines for surveillance will continue to develop as novel germline associations are elucidated, identifiable somatic drivers are discovered, and treatment modalities improve. Future AACR-CPWG meetings will continue to be convened for up-to-date surveillance and intervention recommendations.

Table 1: Childhood Cancer Surveillance for Neurofibromatosis Type I (NF1)

Table 2: Childhood Cancer Surveillance for Noonan syndrome, CBL syndrome, CFC syndrome and NSML

Abbreviations: CFC - Cardiofaciocutaneous syndrome, NSML - Noonan syndrome with multiple lentigines, MPD: myeloproliferative disorder, CBC – complete blood count

Abbreviations: DWI – diffusion weighted imaging, MRI – magnetic resonance imaging, 18F-FDG PET/CT or PET/MRI – 18F-fluorodeoxyglucose positron emission tomography and computed tomography or magnetic resonance imaging, WBMRI – whole-body magnetic resonance imaging

Table 3: Childhood Cancer Surveillance for Costello syndrome

*Malignancies have been reported in CS patients with a PV in *HRAS* predicting p.Gly12 substitutions only. Cancer appears to be less common in rare CS patients with other *HRAS* PVs.

Abbreviations: MRI- magnetic resonance imaging; US- ultrasound; Y- years of age

References

- 1. Hirbe AC, Gutmann DH. Neurofibromatosis type 1: a multidisciplinary approach to care. *Lancet Neurol* 2014;**13**(8):834-43 doi 10.1016/s1474-4422(14)70063-8.
- 2. Kratz CP, Rapisuwon S, Reed H, Hasle H, Rosenberg PS. Cancer in Noonan, Costello, cardiofaciocutaneous and LEOPARD syndromes. *Am J Med Genet C Semin Med Genet* 2011;**157c**(2):83-9 doi 10.1002/ajmg.c.30300.
- 3. Astiazaran-Symonds E, Ney GM, Higgs C, Oba L, Srivastava R, Livinski AA*, et al.* Cancer in Costello syndrome: a systematic review and meta-analysis. *Br J Cancer* 2023;**128**(11):2089-96 doi 10.1038/s41416-023-02229-7.
- 4. Ney G, Gross A, Livinski A, Kratz CP, Stewart DR. Cancer incidence and surveillance strategies in individuals with RASopathies. *Am J Med Genet C Semin Med Genet* 2022;**190**(4):530-40 doi 10.1002/ajmg.c.32018.
- 5. Kratz CP, Franke L, Peters H, Kohlschmidt N, Kazmierczak B, Finckh U*, et al.* Cancer spectrum and frequency among children with Noonan, Costello, and cardio-facio-cutaneous syndromes. *Br J Cancer* 2015;**112**(8):1392-7 doi 10.1038/bjc.2015.75.
- 6. Landry JP, Schertz KL, Chiang YJ, Bhalla AD, Yi M, Keung EZ*, et al.* Comparison of Cancer Prevalence in Patients With Neurofibromatosis Type 1 at an Academic Cancer Center vs in the General Population From 1985 to 2020. *JAMA Netw Open* 2021;**4**(3):e210945 doi 10.1001/jamanetworkopen.2021.0945.
- 7. Dombi E, Baldwin A, Marcus LJ, Fisher MJ, Weiss B, Kim A*, et al.* Activity of Selumetinib in Neurofibromatosis Type 1-Related Plexiform Neurofibromas. *N Engl J Med* 2016;**375**(26):2550- 60 doi 10.1056/NEJMoa1605943.
- 8. Gross AM, Dombi E, Widemann BC. Current status of MEK inhibitors in the treatment of plexiform neurofibromas. *Childs Nerv Syst* 2020;**36**(10):2443-52 doi 10.1007/s00381-020-04731- 2.
- 9. Gross AM, Glassberg B, Wolters PL, Dombi E, Baldwin A, Fisher MJ*, et al.* Selumetinib in children with neurofibromatosis type 1 and asymptomatic inoperable plexiform neurofibroma at risk for developing tumor-related morbidity. *Neuro Oncol* 2022;**24**(11):1978-88 doi 10.1093/neuonc/noac109.
- 10. Miller DT, Freedenberg D, Schorry E, Ullrich NJ, Viskochil D, Korf BR. Health Supervision for Children With Neurofibromatosis Type 1. *Pediatrics* 2019;**143**(5) doi 10.1542/peds.2019-0660.
- 11. Carton C, Evans DG, Blanco I, Friedrich RE, Ferner RE, Farschtschi S*, et al.* ERN GENTURIS tumour surveillance guidelines for individuals with neurofibromatosis type 1. *EClinicalMedicine* 2023;**56**:101818 doi 10.1016/j.eclinm.2022.101818.
- 12. Bergqvist C, Servy A, Valeyrie-Allanore L, Ferkal S, Combemale P, Wolkenstein P. Neurofibromatosis 1 French national guidelines based on an extensive literature review since 1966. *Orphanet J Rare Dis* 2020;**15**(1):37 doi 10.1186/s13023-020-1310-3.
- 13. Evans DGR, Salvador H, Chang VY, Erez A, Voss SD, Schneider KW*, et al.* Cancer and Central Nervous System Tumor Surveillance in Pediatric Neurofibromatosis 1. *Clin Cancer Res* 2017;**23**(12):e46-e53 doi 10.1158/1078-0432.Ccr-17-0589.
- 14. Villani A, Greer MC, Kalish JM, Nakagawara A, Nathanson KL, Pajtler KW*, et al.* Recommendations for Cancer Surveillance in Individuals with RASopathies and Other Rare Genetic Conditions with Increased Cancer Risk. *Clin Cancer Res* 2017;**23**(12):e83-e90 doi 10.1158/1078-0432.Ccr-17-0631.
- 15. Gripp KW, Morse LA, Axelrad M, Chatfield KC, Chidekel A, Dobyns W*, et al.* Costello syndrome: Clinical phenotype, genotype, and management guidelines. *Am J Med Genet A* 2019;**179**(9):1725-44 doi 10.1002/ajmg.a.61270.
- 16. Kallionpää RA, Uusitalo E, Leppävirta J, Pöyhönen M, Peltonen S, Peltonen J. Prevalence of neurofibromatosis type 1 in the Finnish population. *Genetics in Medicine* 2018;**20**(9):1082-6 doi 10.1038/gim.2017.215.
- 17. Friedman JM. Neurofibromatosis 1. In: Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Bean LJH*, et al.*, editors. GeneReviews(®). Seattle (WA): University of Washington, Seattle
- Copyright © 1993-2024, University of Washington, Seattle. GeneReviews is a registered trademark of the University of Washington, Seattle. All rights reserved.; 1993.
- 18. Ejerskov C, Raundahl M, Gregersen PA, Handrup MM. Clinical features and disease severity in patients with mosaic neurofibromatosis type 1: a single-center study and literature review. *Orphanet J Rare Dis* 2021;**16**(1):180 doi 10.1186/s13023-021-01796-3.
- 19. Yang X, Desai K, Agrawal N, Mirchandani K, Chatterjee S, Sarpong E*, et al.* Treatment, Resource Use and Costs Among Pediatric Patients with Neurofibromatosis Type 1 and Plexiform Neurofibromas. *Pediatric Health Med Ther* 2020;**11**:421-8 doi 10.2147/phmt.S265690.
- 20. Legius E, Brems H. Genetic basis of neurofibromatosis type 1 and related conditions, including mosaicism. *Childs Nerv Syst* 2020;**36**(10):2285-95 doi 10.1007/s00381-020-04771-8.
- 21. Bettegowda C, Upadhayaya M, Evans DG, Kim A, Mathios D, Hanemann CO. Genotype-Phenotype Correlations in Neurofibromatosis and Their Potential Clinical Use. *Neurology* 2021;**97**(7 Suppl 1):S91-s8 doi 10.1212/wnl.0000000000012436.
- 22. Cyrus SS, Cohen ASA, Agbahovbe R, Avela K, Yeung KS, Chung BHY*, et al.* Rare SUZ12 variants commonly cause an overgrowth phenotype. *Am J Med Genet C Semin Med Genet* 2019;**181**(4):532-47 doi 10.1002/ajmg.c.31748.
- 23. Legius E, Messiaen L, Wolkenstein P, Pancza P, Avery RA, Berman Y*, et al.* Revised diagnostic criteria for neurofibromatosis type 1 and Legius syndrome: an international consensus recommendation. *Genet Med* 2021;**23**(8):1506-13 doi 10.1038/s41436-021-01170-5.
- 24. National Institutes of Health Consensus Development Conference Statement: neurofibromatosis. Bethesda, Md., USA, July 13-15, 1987. *Neurofibromatosis* 1988;**1**(3):172-8.
- 25. Miettinen MM, Antonescu CR, Fletcher CDM, Kim A, Lazar AJ, Quezado MM*, et al.* Histopathologic evaluation of atypical neurofibromatous tumors and their transformation into malignant peripheral nerve sheath tumor in patients with neurofibromatosis 1-a consensus overview. *Hum Pathol* 2017;**67**:1-10 doi 10.1016/j.humpath.2017.05.010.
- 26. Akshintala S, Baldwin A, Liewehr DJ, Goodwin A, Blakeley JO, Gross AM*, et al.* Longitudinal evaluation of peripheral nerve sheath tumors in neurofibromatosis type 1: growth analysis of plexiform neurofibromas and distinct nodular lesions. *Neuro Oncol* 2020;**22**(9):1368-78 doi 10.1093/neuonc/noaa053.
- 27. Beert E, Brems H, Daniëls B, De Wever I, Van Calenbergh F, Schoenaers J*, et al.* Atypical neurofibromas in neurofibromatosis type 1 are premalignant tumors. *Genes Chromosomes Cancer* 2011;**50**(12):1021-32 doi 10.1002/gcc.20921.
- 28. Fisher MJ, Loguidice M, Gutmann DH, Listernick R, Ferner RE, Ullrich NJ*, et al.* Visual outcomes in children with neurofibromatosis type 1-associated optic pathway glioma following chemotherapy: a multicenter retrospective analysis. *Neuro Oncol* 2012;**14**(6):790-7 doi 10.1093/neuonc/nos076.
- 29. Kotch C, Avery R, Getz KD, Bouffet E, de Blank P, Listernick R*, et al.* Risk factors for treatmentrefractory and relapsed optic pathway glioma in children with neurofibromatosis type 1. *Neuro Oncol* 2022;**24**(8):1377-86 doi 10.1093/neuonc/noac013.
- 30. Fangusaro J, Onar-Thomas A, Poussaint TY, Wu S, Ligon AH, Lindeman N*, et al.* A phase II trial of selumetinib in children with recurrent optic pathway and hypothalamic low-grade glioma without NF1: a Pediatric Brain Tumor Consortium study. *Neuro Oncol* 2021;**23**(10):1777-88 doi 10.1093/neuonc/noab047.
- 31. Fangusaro J, Onar-Thomas A, Young Poussaint T, Wu S, Ligon AH, Lindeman N*, et al.* Selumetinib in paediatric patients with BRAF-aberrant or neurofibromatosis type 1-associated recurrent, refractory, or progressive low-grade glioma: a multicentre, phase 2 trial. *Lancet Oncol* 2019;**20**(7):1011-22 doi 10.1016/s1470-2045(19)30277-3.
- 32. Costa AA, Gutmann DH. Brain tumors in Neurofibromatosis type 1. *Neurooncol Adv* 2019;**1**(1):vdz040 doi 10.1093/noajnl/vdz040.
- 33. Mahdi J, Shah AC, Sato A, Morris SM, McKinstry RC, Listernick R*, et al.* A multi-institutional study of brainstem gliomas in children with neurofibromatosis type 1. *Neurology* 2017;**88**(16):1584-9 doi 10.1212/wnl.0000000000003881.
- 34. Rodriguez EF, Scheithauer BW, Giannini C, Rynearson A, Cen L, Hoesley B*, et al.* PI3K/AKT pathway alterations are associated with clinically aggressive and histologically anaplastic subsets of pilocytic astrocytoma. *Acta Neuropathol* 2011;**121**(3):407-20 doi 10.1007/s00401-010-0784- 9.
- 35. Perreault S, Larouche V, Tabori U, Hawkin C, Lippé S, Ellezam B*, et al.* A phase 2 study of trametinib for patients with pediatric glioma or plexiform neurofibroma with refractory tumor and activation of the MAPK/ERK pathway: TRAM-01. *BMC Cancer* 2019;**19**(1):1250 doi 10.1186/s12885-019-6442-2.
- 36. McCowage GB, Mueller S, Pratilas CA, Hargrave DR, Moertel CL, Whitlock J*, et al.* Trametinib in pediatric patients with neurofibromatosis type 1 (NF-1)–associated plexiform neurofibroma: A phase I/IIa study. *Journal of Clinical Oncology* 2018;**36**(15_suppl):10504- doi 10.1200/JCO.2018.36.15_suppl.10504.
- 37. Fisher MJ, Shih CS, Rhodes SD, Armstrong AE, Wolters PL, Dombi E*, et al.* Cabozantinib for neurofibromatosis type 1-related plexiform neurofibromas: a phase 2 trial. *Nat Med* 2021;**27**(1):165-73 doi 10.1038/s41591-020-01193-6.
- 38. Fasih S, Suppiyah S, Barron J, Barnett-Tapia C, Avery R, Dickson B*, et al.* Malignant transformation of plexiform neurofibroma to MPNST while on MEK inhibitor. *Neurooncol Adv* 2021;**3**(1):vdab033 doi 10.1093/noajnl/vdab033.
- 39. Higham CS, Dombi E, Rogiers A, Bhaumik S, Pans S, Connor SEJ*, et al.* The characteristics of 76 atypical neurofibromas as precursors to neurofibromatosis 1 associated malignant peripheral nerve sheath tumors. *Neuro Oncol* 2018;**20**(6):818-25 doi 10.1093/neuonc/noy013.
- 40. Pemov A, Hansen NF, Sindiri S, Patidar R, Higham CS, Dombi E*, et al.* Low mutation burden and frequent loss of CDKN2A/B and SMARCA2, but not PRC2, define premalignant neurofibromatosis type 1-associated atypical neurofibromas. *Neuro Oncol* 2019;**21**(8):981-92 doi 10.1093/neuonc/noz028.
- 41. Nelson CN, Dombi E, Rosenblum JS, Miettinen MM, Lehky TJ, Whitcomb PO*, et al.* Safe marginal resection of atypical neurofibromas in neurofibromatosis type 1. *J Neurosurg* 2019:1-11 doi 10.3171/2019.7.Jns191353.
- 42. Bernthal NM, Putnam A, Jones KB, Viskochil D, Randall RL. The effect of surgical margins on outcomes for low grade MPNSTs and atypical neurofibroma. *J Surg Oncol* 2014;**110**(7):813-6 doi 10.1002/jso.23736.
- 43. Vaassen P, Feldkamp A, Scholz M, Blau T, Dürr NR, Rosenbaum T. A chance to cut is a chance to cure: complete resection of an atypical neurofibroma prevents further progression to malignancy. *Childs Nerv Syst* 2023;**39**(11):3301-4 doi 10.1007/s00381-023-06029-5.
- 44. De Raedt T, Brems H, Wolkenstein P, Vidaud D, Pilotti S, Perrone F*, et al.* Elevated risk for MPNST in NF1 microdeletion patients. *Am J Hum Genet* 2003;**72**(5):1288-92 doi 10.1086/374821.
- 45. Koczkowska M, Chen Y, Callens T, Gomes A, Sharp A, Johnson S*, et al.* Genotype-Phenotype Correlation in NF1: Evidence for a More Severe Phenotype Associated with Missense Mutations Affecting NF1 Codons 844-848. *Am J Hum Genet* 2018;**102**(1):69-87 doi 10.1016/j.ajhg.2017.12.001.
- 46. Nguyen R, Jett K, Harris GJ, Cai W, Friedman JM, Mautner VF. Benign whole body tumor volume is a risk factor for malignant peripheral nerve sheath tumors in neurofibromatosis type 1. *J Neurooncol* 2014;**116**(2):307-13 doi 10.1007/s11060-013-1293-1.
- 47. Evans DG, Birch JM, Ramsden RT, Sharif S, Baser ME. Malignant transformation and new primary tumours after therapeutic radiation for benign disease: substantial risks in certain tumour prone syndromes. *J Med Genet* 2006;**43**(4):289-94 doi 10.1136/jmg.2005.036319.
- 48. Yamanaka R, Hayano A. Radiation-Induced Malignant Peripheral Nerve Sheath Tumors: A Systematic Review. *World Neurosurg* 2017;**105**:961-70.e8 doi 10.1016/j.wneu.2017.06.010.
- 49. Malbari F, Spira M, P BK, Zhu C, Roth M, Gill J*, et al.* Malignant Peripheral Nerve Sheath Tumors in Neurofibromatosis: Impact of Family History. *J Pediatr Hematol Oncol* 2018;**40**(6):e359-e63 doi 10.1097/mph.0000000000001156.
- 50. Prudner BC, Ball T, Rathore R, Hirbe AC. Diagnosis and management of malignant peripheral nerve sheath tumors: Current practice and future perspectives. *Neurooncol Adv* 2020;**2**(Suppl 1):i40-i9 doi 10.1093/noajnl/vdz047.
- 51. McGaughran JM, Harris DI, Donnai D, Teare D, MacLeod R, Westerbeek R*, et al.* A clinical study of type 1 neurofibromatosis in north west England. *J Med Genet* 1999;**36**(3):197-203.
- 52. Patil S, Chamberlain RS. Neoplasms associated with germline and somatic NF1 gene mutations. *Oncologist* 2012;**17**(1):101-16 doi 10.1634/theoncologist.2010-0181.
- 53. Crucis A, Richer W, Brugières L, Bergeron C, Marie-Cardine A, Stephan JL*, et al.* Rhabdomyosarcomas in children with neurofibromatosis type I: A national historical cohort. *Pediatr Blood Cancer* 2015;**62**(10):1733-8 doi 10.1002/pbc.25556.
- 54. Li H, Sisoudiya SD, Martin-Giacalone BA, Khayat MM, Dugan-Perez S, Marquez-Do DA*, et al.* Germline Cancer Predisposition Variants in Pediatric Rhabdomyosarcoma: A Report From the Children's Oncology Group. *J Natl Cancer Inst* 2021;**113**(7):875-83 doi 10.1093/jnci/djaa204.
- 55. Stewart DR, Korf BR, Nathanson KL, Stevenson DA, Yohay K. Care of adults with neurofibromatosis type 1: a clinical practice resource of the American College of Medical Genetics and Genomics (ACMG). *Genet Med* 2018;**20**(7):671-82 doi 10.1038/gim.2018.28.
- 56. Uusitalo E, Kallionpää RA, Kurki S, Rantanen M, Pitkäniemi J, Kronqvist P*, et al.* Breast cancer in neurofibromatosis type 1: overrepresentation of unfavourable prognostic factors. *Br J Cancer* 2017;**116**(2):211-7 doi 10.1038/bjc.2016.403.
- 57. Seminog OO, Goldacre MJ. Age-specific risk of breast cancer in women with neurofibromatosis type 1. *Br J Cancer* 2015;**112**(9):1546-8 doi 10.1038/bjc.2015.78.
- 58. Wilding M, Fleming J, Moore K, Crook A, Reddy R, Choi S*, et al.* Clinical and imaging modality factors impacting radiological interpretation of breast screening in young women with neurofibromatosis type 1. *Fam Cancer* 2023;**22**(4):499-511 doi 10.1007/s10689-023-00340-5.
- 59. Uusitalo E, Rantanen M, Kallionpää RA, Pöyhönen M, Leppävirta J, Ylä-Outinen H*, et al.* Distinctive Cancer Associations in Patients With Neurofibromatosis Type 1. *J Clin Oncol* 2016;**34**(17):1978-86 doi 10.1200/jco.2015.65.3576.
- 60. Stewart DR, Sloan JL, Yao L, Mannes AJ, Moshyedi A, Lee CC*, et al.* Diagnosis, management, and complications of glomus tumours of the digits in neurofibromatosis type 1. *J Med Genet* 2010;**47**(8):525-32 doi 10.1136/jmg.2009.073965.
- 61. de Blank PMK, Fisher MJ, Liu GT, Gutmann DH, Listernick R, Ferner RE*, et al.* Optic Pathway Gliomas in Neurofibromatosis Type 1: An Update: Surveillance, Treatment Indications, and Biomarkers of Vision. *J Neuroophthalmol* 2017;**37 Suppl 1**(Suppl 1):S23-s32 doi 10.1097/wno.0000000000000550.
- 62. Tang Y, Gutmann DH. Neurofibromatosis Type 1-Associated Optic Pathway Gliomas: Current Challenges and Future Prospects. *Cancer Manag Res* 2023;**15**:667-81 doi 10.2147/cmar.S362678.
- 63. Prada CE, Hufnagel RB, Hummel TR, Lovell AM, Hopkin RJ, Saal HM*, et al.* The Use of Magnetic Resonance Imaging Screening for Optic Pathway Gliomas in Children with Neurofibromatosis Type 1. *J Pediatr* 2015;**167**(4):851-6.e1 doi 10.1016/j.jpeds.2015.07.001.
- 64. Brenner W, Friedrich RE, Gawad KA, Hagel C, von Deimling A, de Wit M*, et al.* Prognostic relevance of FDG PET in patients with neurofibromatosis type-1 and malignant peripheral nerve sheath tumours. *Eur J Nucl Med Mol Imaging* 2006;**33**(4):428-32 doi 10.1007/s00259-005-0030- 1.
- 65. Dombi E, Solomon J, Gillespie AJ, Fox E, Balis FM, Patronas N*, et al.* NF1 plexiform neurofibroma growth rate by volumetric MRI: relationship to age and body weight. *Neurology* 2007;**68**(9):643- 7 doi 10.1212/01.wnl.0000250332.89420.e6.
- 66. Ahlawat S, Fayad LM, Khan MS, Bredella MA, Harris GJ, Evans DG*, et al.* Current whole-body MRI applications in the neurofibromatoses: NF1, NF2, and schwannomatosis. *Neurology* 2016;**87**(7 Suppl 1):S31-9 doi 10.1212/wnl.0000000000002929.
- 67. Fisher MJ, Blakeley JO, Weiss BD, Dombi E, Ahlawat S, Akshintala S*, et al.* Management of neurofibromatosis type 1-associated plexiform neurofibromas. *Neuro Oncol* 2022;**24**(11):1827- 44 doi 10.1093/neuonc/noac146.
- 68. Mautner VF, Asuagbor FA, Dombi E, Fünsterer C, Kluwe L, Wenzel R*, et al.* Assessment of benign tumor burden by whole-body MRI in patients with neurofibromatosis 1. *Neuro Oncol* 2008;**10**(4):593-8 doi 10.1215/15228517-2008-011.
- 69. Szymanski JJ, Sundby RT, Jones PA, Srihari D, Earland N, Harris PK*, et al.* Cell-free DNA ultra-lowpass whole genome sequencing to distinguish malignant peripheral nerve sheath tumor (MPNST) from its benign precursor lesion: A cross-sectional study. *PLoS Med* 2021;**18**(8):e1003734 doi 10.1371/journal.pmed.1003734.
- 70. Cortes-Ciriano I, Steele CD, Piculell K, Al-Ibraheemi A, Eulo V, Bui MM*, et al.* Genomic Patterns of Malignant Peripheral Nerve Sheath Tumor (MPNST) Evolution Correlate with Clinical Outcome and Are Detectable in Cell-Free DNA. *Cancer Discov* 2023;**13**(3):654-71 doi 10.1158/2159- 8290.Cd-22-0786.
- 71. Tartaglia M, Mehler EL, Goldberg R, Zampino G, Brunner HG, Kremer H*, et al.* Mutations in PTPN11, encoding the protein tyrosine phosphatase SHP-2, cause Noonan syndrome. *Nat Genet* 2001;**29**(4):465-8 doi 10.1038/ng772.
- 72. Roberts AE, Allanson JE, Tartaglia M, Gelb BD. Noonan syndrome. *Lancet* 2013;**381**(9863):333- 42 doi 10.1016/s0140-6736(12)61023-x.
- 73. Gross AM, Frone M, Gripp KW, Gelb BD, Schoyer L, Schill L*, et al.* Advancing RAS/RASopathy therapies: An NCI-sponsored intramural and extramural collaboration for the study of RASopathies. *Am J Med Genet A* 2020;**182**(4):866-76 doi 10.1002/ajmg.a.61485.
- 74. Zenker M, Edouard T, Blair JC, Cappa M. Noonan syndrome: improving recognition and diagnosis. *Arch Dis Child* 2022;**107**(12):1073-8 doi 10.1136/archdischild-2021-322858.
- 75. Motta M, Fasano G, Gredy S, Brinkmann J, Bonnard AA, Simsek-Kiper PO*, et al.* SPRED2 loss-offunction causes a recessive Noonan syndrome-like phenotype. *Am J Hum Genet* 2021;**108**(11):2112-29 doi 10.1016/j.ajhg.2021.09.007.
- 76. Onore ME, Caiazza M, Farina A, Scarano G, Budillon A, Borrelli RN*, et al.* A Novel Homozygous Loss-of-Function Variant in SPRED2 Causes Autosomal Recessive Noonan-like Syndrome. *Genes (Basel)* 2023;**15**(1) doi 10.3390/genes15010032.
- 77. Roberts AE. Noonan Syndrome. In: Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Bean LJH*, et al.*, editors. GeneReviews(®). Seattle (WA): University of Washington, Seattle
- Copyright © 1993-2024, University of Washington, Seattle. GeneReviews is a registered trademark of the University of Washington, Seattle. All rights reserved.; 1993.
- 78. Strullu M, Caye A, Lachenaud J, Cassinat B, Gazal S, Fenneteau O*, et al.* Juvenile myelomonocytic leukaemia and Noonan syndrome. *J Med Genet* 2014;**51**(10):689-97 doi 10.1136/jmedgenet-2014-102611.
- 79. Alhumaid MS, Dasouki MJ, Ahmed SO, AbalKhail H, Hagos S, Wakil S*, et al.* Comprehensive Genomic Analysis of Noonan Syndrome and Acute Myeloid Leukemia in Adults: A Review and Future Directions. *Acta Haematol* 2020;**143**(6):583-93 doi 10.1159/000505715.
- 80. Yang F, Long N, Anekpuritanang T, Bottomly D, Savage JC, Lee T*, et al.* Identification and prioritization of myeloid malignancy germline variants in a large cohort of adult patients with AML. *Blood* 2022;**139**(8):1208-21 doi 10.1182/blood.2021011354.
- 81. Siegfried A, Cances C, Denuelle M, Loukh N, Tauber M, Cavé H*, et al.* Noonan syndrome, PTPN11 mutations, and brain tumors. A clinical report and review of the literature. *Am J Med Genet A* 2017;**173**(4):1061-5 doi 10.1002/ajmg.a.38108.
- 82. Gupta AK, Meena JP, Chopra A, Tanwar P, Seth R. Juvenile myelomonocytic leukemia-A comprehensive review and recent advances in management. *Am J Blood Res* 2021;**11**(1):1-21.
- 83. Niihori T, Aoki Y, Ohashi H, Kurosawa K, Kondoh T, Ishikiriyama S*, et al.* Functional analysis of PTPN11/SHP-2 mutants identified in Noonan syndrome and childhood leukemia. *J Hum Genet* 2005;**50**(4):192-202 doi 10.1007/s10038-005-0239-7.
- 84. Hofmans M, Schröder R, Lammens T, Flotho C, Niemeyer C, Van Roy N*, et al.* Noonan syndromeassociated myeloproliferative disorder with somatically acquired monosomy 7: impact on clinical decision making. *Br J Haematol* 2019;**187**(4):E83-e6 doi 10.1111/bjh.16191.
- 85. Niemeyer CM, Kang MW, Shin DH, Furlan I, Erlacher M, Bunin NJ*, et al.* Germline CBL mutations cause developmental abnormalities and predispose to juvenile myelomonocytic leukemia. *Nat Genet* 2010;**42**(9):794-800 doi 10.1038/ng.641.
- 86. Pérez B, Mechinaud F, Galambrun C, Ben Romdhane N, Isidor B, Philip N*, et al.* Germline mutations of the CBL gene define a new genetic syndrome with predisposition to juvenile myelomonocytic leukaemia. *J Med Genet* 2010;**47**(10):686-91 doi 10.1136/jmg.2010.076836.
- 87. Mayerhofer C, Niemeyer CM, Flotho C. Current Treatment of Juvenile Myelomonocytic Leukemia. *J Clin Med* 2021;**10**(14) doi 10.3390/jcm10143084.
- 88. Giannoulatou E, McVean G, Taylor IB, McGowan SJ, Maher GJ, Iqbal Z*, et al.* Contributions of intrinsic mutation rate and selfish selection to levels of de novo HRAS mutations in the paternal germline. *Proc Natl Acad Sci U S A* 2013;**110**(50):20152-7 doi 10.1073/pnas.1311381110.
- 89. Gripp KW, Scott CI, Jr., Nicholson L, McDonald-McGinn DM, Ozeran JD, Jones MC*, et al.* Five additional Costello syndrome patients with rhabdomyosarcoma: proposal for a tumor screening protocol. *Am J Med Genet* 2002;**108**(1):80-7 doi 10.1002/ajmg.10241.
- 90. Leoni C, Viscogliosi G, Tartaglia M, Aoki Y, Zampino G. Multidisciplinary Management of Costello Syndrome: Current Perspectives. *J Multidiscip Healthc* 2022;**15**:1277-96 doi 10.2147/jmdh.S291757.
- 91. Robbins KM, Stabley DL, Holbrook J, Sahraoui R, Sadreameli A, Conard K*, et al.* Paternal uniparental disomy with segmental loss of heterozygosity of chromosome 11 are hallmark characteristics of syndromic and sporadic embryonal rhabdomyosarcoma. *Am J Med Genet A* 2016;**170**(12):3197-206 doi 10.1002/ajmg.a.37949.
- 92. Casanova M, Meazza C, Favini F, Fiore M, Morosi C, Ferrari A. Rhabdomyosarcoma of the extremities: a focus on tumors arising in the hand and foot. *Pediatr Hematol Oncol* 2009;**26**(5):321-31 doi 10.1080/08880010902964367.
- 93. Kamihara J, Diller LR, Foulkes WD, Michaeli O, Nakano Y, Pajtler KW*, et al.* Neuroblastoma Predisposition and Surveillance-An Update from the 2023 AACR Childhood Cancer Predisposition Workshop. *Clin Cancer Res* 2024:Of1-of7 doi 10.1158/1078-0432.Ccr-24-0237.
- 94. Gripp KW, Kawame H, Viskochil DH, Nicholson L. Elevated catecholamine metabolites in patients with Costello syndrome. *Am J Med Genet A* 2004;**128a**(1):48-51 doi 10.1002/ajmg.a.30100.
- 95. Leoni C, Paradiso FV, Foschi N, Tedesco M, Pierconti F, Silvaroli S*, et al.* Prevalence of bladder cancer in Costello syndrome: New insights to drive clinical decision-making. *Clin Genet* 2022;**101**(4):454-8 doi 10.1111/cge.14111.
- 96. Tartaglia M, Martinelli S, Stella L, Bocchinfuso G, Flex E, Cordeddu V*, et al.* Diversity and functional consequences of germline and somatic PTPN11 mutations in human disease. *Am J Hum Genet* 2006;**78**(2):279-90 doi 10.1086/499925.
- 97. Pandit B, Sarkozy A, Pennacchio LA, Carta C, Oishi K, Martinelli S*, et al.* Gain-of-function RAF1 mutations cause Noonan and LEOPARD syndromes with hypertrophic cardiomyopathy. *Nat Genet* 2007;**39**(8):1007-12 doi 10.1038/ng2073.
- 98. Nishi E, Mizuno S, Nanjo Y, Niihori T, Fukushima Y, Matsubara Y*, et al.* A novel heterozygous MAP2K1 mutation in a patient with Noonan syndrome with multiple lentigines. *Am J Med Genet A* 2015;**167a**(2):407-11 doi 10.1002/ajmg.a.36842.
- 99. Koudova M, Seemanova E, Zenker M. Novel BRAF mutation in a patient with LEOPARD syndrome and normal intelligence. *Eur J Med Genet* 2009;**52**(5):337-40 doi 10.1016/j.ejmg.2009.04.006.
- 100. Sarkozy A, Carta C, Moretti S, Zampino G, Digilio MC, Pantaleoni F*, et al.* Germline BRAF mutations in Noonan, LEOPARD, and cardiofaciocutaneous syndromes: molecular diversity and associated phenotypic spectrum. *Hum Mutat* 2009;**30**(4):695-702 doi 10.1002/humu.20955.
- 101. Gelb BD, Tartaglia M. Noonan Syndrome with Multiple Lentigines. In: Adam MP, Feldman J, Mirzaa GM, Pagon RA, Wallace SE, Bean LJH*, et al.*, editors. GeneReviews(®). Seattle (WA): University of Washington, Seattle
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- 102. Palacios-Diaz RD, Pozuelo-Ruiz M, De Unamuno-Bustos B, Llavador-Ros M, Botella-Estrada R. Melanoma and LEOPARD Syndrome: Understanding the Role of PTPN11 Mutations in Melanomagenesis. *Acta Derm Venereol* 2024;**104**:adv14720 doi 10.2340/actadv.v104.14720.
- 103. Scorrano G, David E, Calì E, Chimenz R, La Bella S, Di Ludovico A*, et al.* The Cardiofaciocutaneous Syndrome: From Genetics to Prognostic-Therapeutic Implications. *Genes (Basel)* 2023;**14**(12) doi 10.3390/genes14122111.
- 104. Pierpont ME, Magoulas PL, Adi S, Kavamura MI, Neri G, Noonan J*, et al.* Cardio-facio-cutaneous syndrome: clinical features, diagnosis, and management guidelines. *Pediatrics* 2014;**134**(4):e1149-62 doi 10.1542/peds.2013-3189.
- 105. Brems H, Chmara M, Sahbatou M, Denayer E, Taniguchi K, Kato R*, et al.* Germline loss-offunction mutations in SPRED1 cause a neurofibromatosis 1-like phenotype. *Nat Genet* 2007;**39**(9):1120-6 doi 10.1038/ng2113.
- 106. Denayer E, Legius E. Legius Syndrome and its Relationship with Neurofibromatosis Type 1. *Acta Derm Venereol* 2020;**100**(7):adv00093 doi 10.2340/00015555-3429.
- 107. Pasmant E, Ballerini P, Lapillonne H, Perot C, Vidaud D, Leverger G*, et al.* SPRED1 disorder and predisposition to leukemia in children. *Blood* 2009;**114**(5):1131 doi 10.1182/blood-2009-04- 218503.
- 108. Pasmant E, Gilbert-Dussardier B, Petit A, de Laval B, Luscan A, Gruber A*, et al.* SPRED1, a RAS MAPK pathway inhibitor that causes Legius syndrome, is a tumour suppressor downregulated in paediatric acute myeloblastic leukaemia. *Oncogene* 2015;**34**(5):631-8 doi 10.1038/onc.2013.587.